ORIGINAL ARTICLE

Reactivation of acetycholinesterase inhibited by the pesticide chlorpyrifos

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Summary

Organophosphorus pesticides such as parathion or chlorpyrifos are substances used worldwide for agricultural purposes. These compounds are able to inhibit an enzyme acetylcholinesterase (AChE; EC 3.1.1.7) by phosphorylation in its active site. AChE reactivators and anticholinergics are generally used as antidotes in the case of intoxication by these agents. In this work, the reactivation potency of nine structurally different AChE reactivators was tested *in vitro*. Chlorpyrifos was chosen as an appropriate member of the pesticide family. The result is that bisquaternary reactivators with two oxime groups in position four at the pyridinium rings (trimedoxime and K074) seem to be the most potent reactivators of chlorpyrifos-inhibited AChE.

Keywords: acetylcholinesterase - reactivator - pesticide - organophosphate

INTRODUCTION

Self-poisoning with pesticides is a major problem across the world. It is estimated that hundreds of thousands of people die each year due to the into-

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xication by organophosphorus pesticides (Gunnell and Eddleston, 2003; Meggs 2003). Although these compounds act in the same way as nerve agents [inhibition of the enzyme acetylcholinesterase (AChE; EC 3.1.1.7)], nerve agent antidotes are not generally as satisfactory in the treatment of pesticide poisoning as in the case of nerve agent poisoning (Worek et al. 1996, 1999). This is due to the fact that pesticides create a different enzymeinhibitor complex than with nerve agents. Pesticides create the dimethoxy-(methylchlorpyrifos, fenitrothion, trichlorfon), diethoxy- (chlorpyrifos, parathion, paraoxon) or diisopropoxy- (DFP) phosphoryl complex with enzyme (Figure 1). As mentioned above, these complexes are poorly reactivated by the commonly used AChE reactivators. Of the commercially

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available oximes, only obidoxime seems to be satisfactorily effective in the case of pesticide poisonings (Thiermann et al. 1997, Musílek et al. 2005). However, it depends on the pesticide used and on the time of therapy administration (Thiermann et al. 1997, Eddleston et al. 2005).

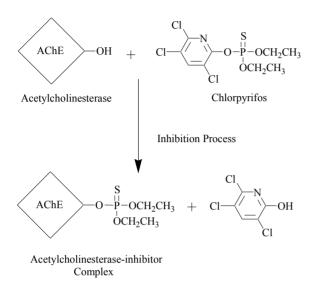


Fig.1. Inhibition of acetylcholinesterase by the pesticide chlorpyrifos.

Currently, there is a lack of the information necessary for the successful development of antidotes against pesticide poisoning (Buckley et al. 2005). Because of this, we would like to focus on the relationship between the structure of cholinesterase reactivators and their biological activity. This study could be helpful in the future to scientists working on the development of new antidotes.

To obtain the basic structural requirements of AChE reactivators for the treatment of pesticide poisonings, we have selected nine known AChE reactivators differing in their chemical structure. Three compounds tested were monoquaternary AChE reactivators differing in the position of the oxime group (position 2, 3 or 4 on the pyridinium ring). Among them, pralidoxime, an oxime currently available for first aid in the US, is included. The remaining six AChE reactivators were bisquaternary, differing in the position of the oxime groups on the pyridinium rings (position 2, 3) or 4) and in the length of the connection chain (three or four membered linker) between both pyridinium rings. As described earlier, all these structural factors (the number of quaternary nitrogens, number and position of oxime groups, and length of the connection chain) play a very important role in the biological activity of AChE reactivators (Kuča et al. 2006).

In this work, all the above-mentioned reactivators were tested *in vitro* as potential reactivators of AChE inhibited by pesticide chlorpyrifos.

MATERIAL AND METHODS

All tested AChE reactivators were synthesized in our department (Musílek et al. 2006). Their purity was tested by TLC using DC-Alufolien Cellulose F (Merck, Germany) and elution BuOH-CH₃COOH-H₂O 5: 1: 2, detection by solution of Dragendorf's reagent (solution containing 10 ml CH₃COOH, 50 ml H₂O and 5 cm³ of basic solution prepared by mixing of two fractions - fraction I.: 0.85 mg $Bi(NO_3)_3$, 40 cm³ H₂O, 10 cm³ CH₃COOH; fraction II.: 8 g KI, 20 cm³ H₂O). The pesticide chlorpyrifos (O.O-diethyl-O-(3,5,6-trichloropyrid-2-yl)phosphorothioate) was obtained from Riedel-de Haen of analytical standard with 99.2 % purity. All other chemicals used in this experiment were purchased from Fluka and Aldrich (Czech Republic) and used without further purification.

The in vitro testing of the synthesized reactivators involved the standard collection of experimental procedures. The whole method is described in detail in the work of Kuča and Cabal (2005). The reactivation efficacy of tested reactivators was evaluated in 10% rat brain homogenate that was incubated with chlorpyrifos for 30 minutes and then the tested oxime of appropriate concentration $(10^{-5} \text{ and } 10^{-3} \text{ M})$ was added. After 10 min of reactivation, the activity of brain AChE was measured using the potentiostatic method with an automatic titrator RTS 822 (Radiometer, Denmark). The data on the initial rate of enzyme reaction with the substrate made it possible to calculate the percentage of increase in the activity of reactivated enzyme in the reaction mixture.

RESULTS

All results obtained are shown in Fig. 2 and Table 1. As can be seen clearly, all the tested oximes were able to reactivate chlorpyrifos-inhibited AChE. Six oximes reached a reactivation potency of over 10%, which is believed to be sufficient for the survival of intoxicated rats. The highest reactivation potency was achieved for pralidoxime (80%) and obidoxime (79%). Unfortunately, this reactivation potency was not reached at the concentration 10^{-5} M sufficient for humans (Bajgar 2004). At relevant doses for

humans, trimedoxime (33%) and K074 (30%) seem to be the most potent reactivators tested.

have in their structure two quaternary nitrogens, connected with three or four membered linking chain. Moreover, both reactivators have two oxime groups in position four at two pyridinium rings.

If we focus on structural factors, both the most potent AChE reactivators (trimedoxime and K074)

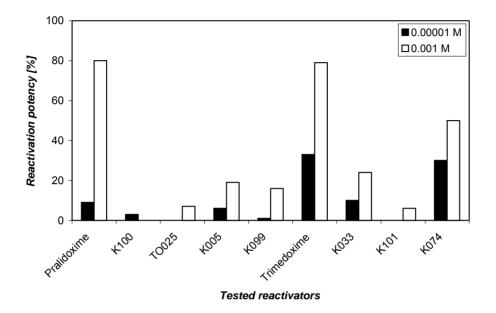


Fig. 2. The reactivation potency of the tested acetylcholinesterase reactivators (Source of enzyme – rat brain homogenate; inhibitor – chlorpyrifos; time of inhibition – 30 min; time of reactivation – 10 min; pH 7.6; 25 °C).

DISCUSSION

Owing to the fact that all the earlier developed AChE reactivators were prepared with the aim of treating nerve agent intoxication, there is a lack of reactivators for treatment of pesticide poisonings, because of the low efficacy of nerve agentreactivators in the case of pesticide intoxications. It is generally known that oxime HI-6, so promising in the case of sarin, cyclosarin and VX poisoning, is practically ineffective in the case of pesticide reactivation (Worek et al. 1996, 1999, Musílek et al. 2005). Currently, obidoxime seems to be the reactivator of the first choice for the treatment of pesticide-intoxication (Thiermann 1997). Its structural analogue, trimedoxime, achieved according to our former results similar reactivation potency (Musilek et al. 2005). However due to the potential toxicity of both reactivators, these substances should not be used over the long period of treatment necessary for achieving satisfactory cholinesterase activity in blood (Bismuth et al. 1992, Balali-Mood and Shariat 1998, Jun et al., 2006).

Therefore the development of new more potent and less toxic antidotes, especially AChE reactivators, for the treatment of pesticide poisonings should continue. To design a new, more potent AChE reactivator, it would be very important to know the relationship between the structure of reactivators and their biological activity (Gray 1984, Kuča et al. 2006). This was a goal in this work.

As is known from our previous results, there are several structural factors (the number of quaternary nitrogens, number and position of oxime groups, and the length of the connection chain) influencing the activity of currently used nerve agent-inhibited AChE reactivators (Kuča et al. 2003, Kuča et al. 2006). We wanted to find out if these factors influence the reactivation potency of the tested oximes in the same manner as they did in the case of nerve agents.

It transpired that bisquaternary are more potent than monoquaternary reactivators. The same results were obtained also for nerve agents-inhibited AChE. Secondly, the ideal position for the oxime group seems to be position four. This result is in good correlation with reactivators of tabuninhibited AChE (Cabal et al. 2004). The position of the oxime group is different in cyclosarin-inhibited AChE reactivators, which have the oxime group in position two at the pyridinium ring (except methoxime – due to its size) (Kassa and Cabal 1999, Kuča and Patočka 2004). In this manuscript, we could not discuss the dependence of the reactivation potency of AChE reactivators on the length of linker, because in the study we concentrated only three and four membered connection chains, which are believed to be the best for the reactivation of nerve agent inhibited AChE (Kuča et al. 2003).

The aim of our work was to focus the attention of scientists working in this field on the targeted development of new pesticide-inhibited AChE reactivators and, moreover, to point out that there are several structural factors, which should be followed in producing new AChE reactivators with a promising reactivation potency

Table 1. The reactivation potency and structural factors of tested reactivators (Source of enzyme – rat brain homogenate; inhibitor – chlorpyrifos; time of inhibition – 30 min; time of reactivation – 10 min; pH 7.6; 25°C)

Structures		Reactivation Potency [%]		Structural Factors		
	Reactivator	0.00001 M	0.001 M	Oxime group position	Length of the connection chain	Number of quaternary nitrogens
ЛОН	Pralidoxime	9	80	2	_	1
	K100	3	0	3	-	1
Ň	TO 025	0	7	4	-	1
CH ₃ I						
/NOI	H					
$\left[\frac{1}{1}, \frac{1}{2} \right]$	K005	6	19	2.2′	3	2
	K099	1	16	3.3′	3	2
2Br ⁻ (CH.)	Trimedoxime	33	79	4.4′	3	2
^{2Br} (CH ₂) _n	K033	10	24	2.2′	4	2
/==N	K101	0	6	3.3′	4	2
+ NOH	K074	30	50	4.4′	4	2

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